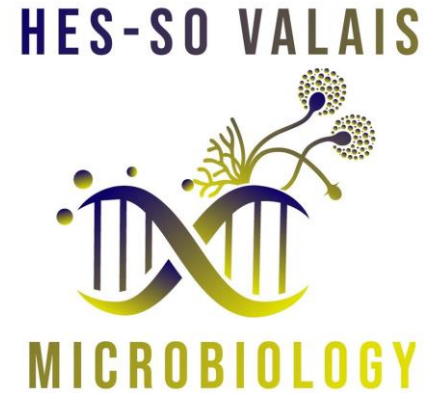


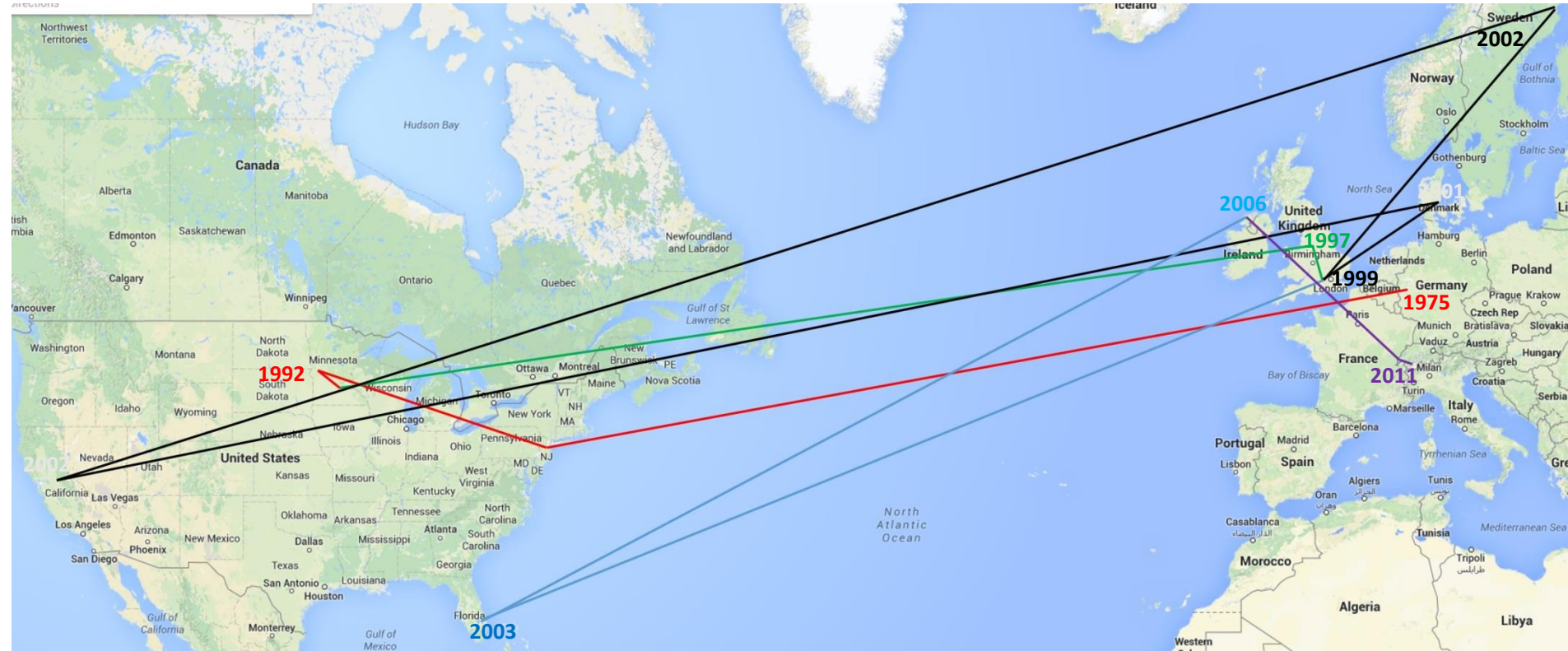
hes.
so
you.

Sustainable materials from food wastes



Dr. Wolfram Brück

<https://people.hes-so.ch/de/profile/wolfram.bruck>
wolfram.bruck@hevs.ch



Employment history

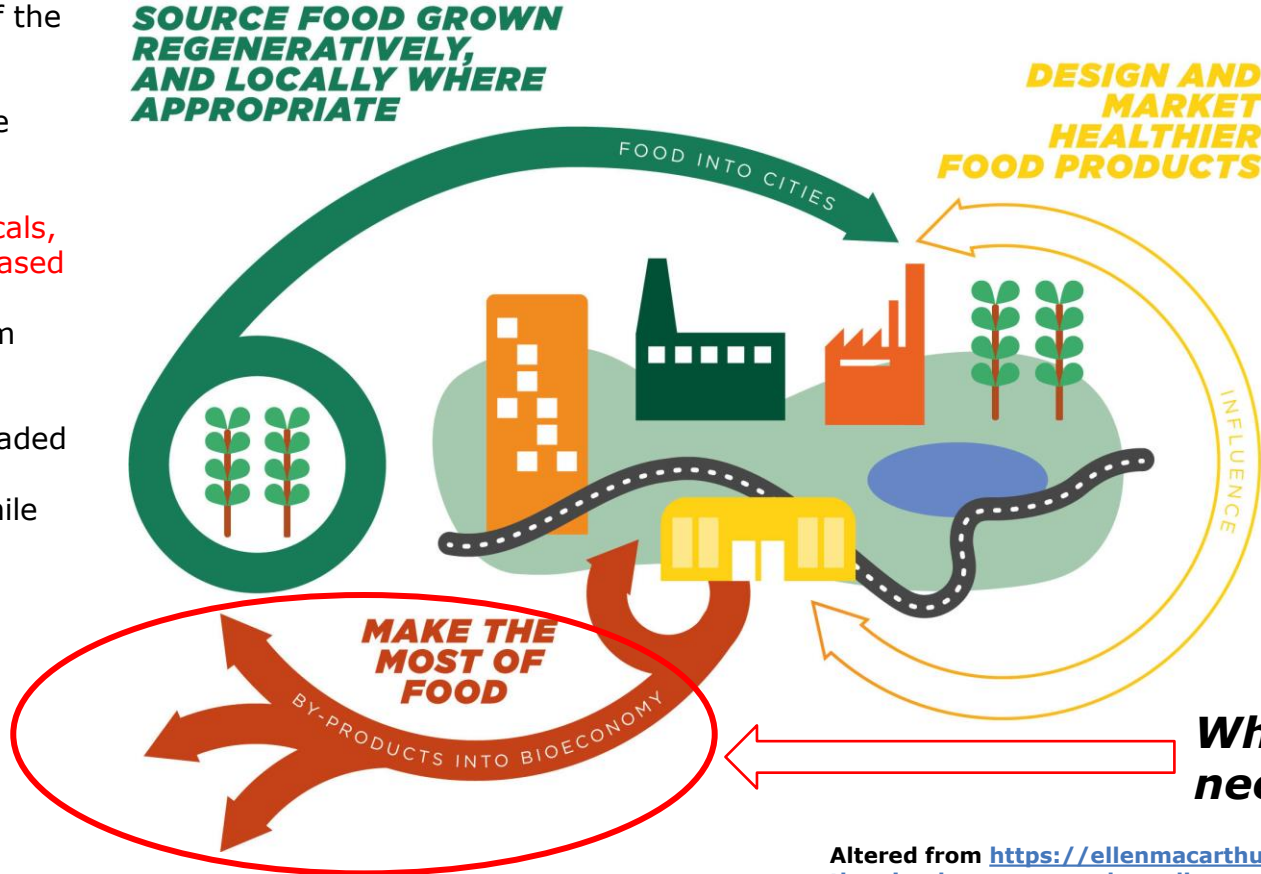
- 1995 – 1997 B.S. Microbiology, St. Cloud State University, MN, US
- 1997 – 1998 M.Sc. Medical and Molecular Microbiology, Univ. Manchester Medical School, UK
- 1999 – 2003 Ph.D. Food Microbiology (infant gut health) University of Reading, UK (supervisor Prof. Dr. Glenn Gibson)
- 2003 – 2005 Postdoc at Harbor Branch Oceanographic Inst. (Florida) Marine Microbial Ecology
- 2006 – 2011 Director of the Center of Applied Marine Microbiology, Marine Microbial Ecology (Co. Donegal, Ireland)
- 2011 – 2014 R&D Specialist at Nestlé Research Center (follow-on milk, probiotics & postbiotics), BSO
- 2014 – current Professor for microbiology and foodomics at the HES-SO Valais - Head of Food Safety Laboratory (ISO17025 and GMP), BSO





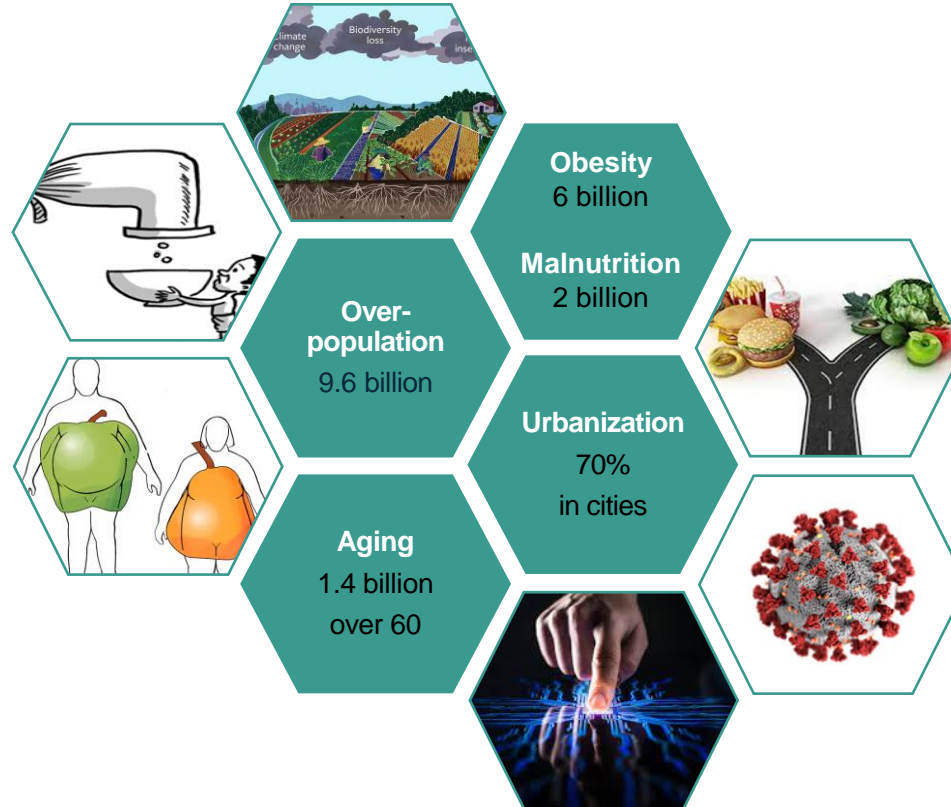
A circular food bioeconomy

«The concept of the **circular bioeconomy** is described as the production of energy, food, platform chemicals, and other bio-based materials and compounds from biomass in a sustainable and integrated/cascaded manner (biorefinery) while generating **zero waste**»⁴



Where we need to act!

Relevant global drivers – a vision to 2050



> The integrated biowaste biorefinery approach at HES-SO Valais/Wallis (Microbiology laboratories)



Oligosaccharides

Products:
Prebiotics
Thickeners
Laxatives
Stabilizers

Proteins

Products:
Meat analogs
Dairy analogs
Enzymes
Peptide prebiotics

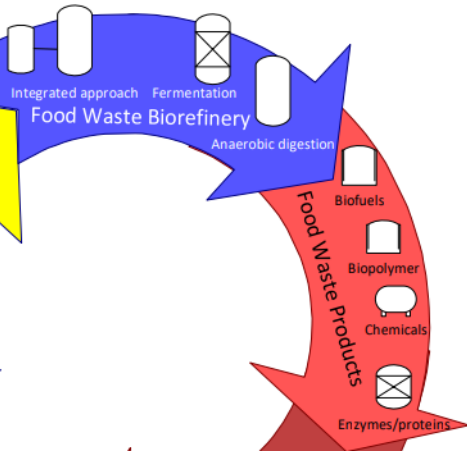
Polyphenols

Products:
Functional food additives

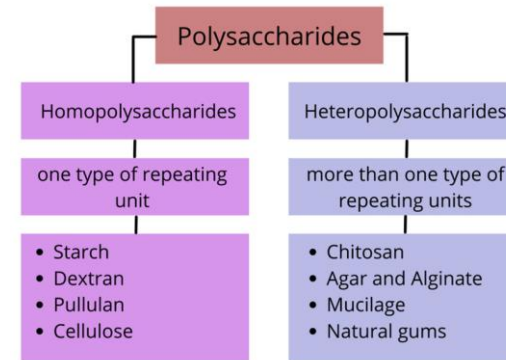
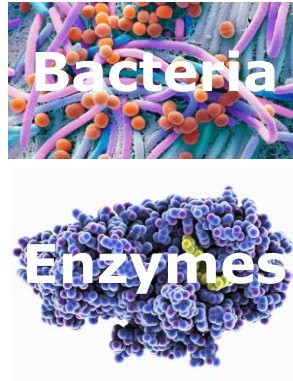
Base chemicals

Products:
Food packaging
Edible films
Antimicrobial barriers

Available key technology and knowledge base: Microbial fermentation, anaerobic digestion, algae production systems, ex vivo models, foodomics



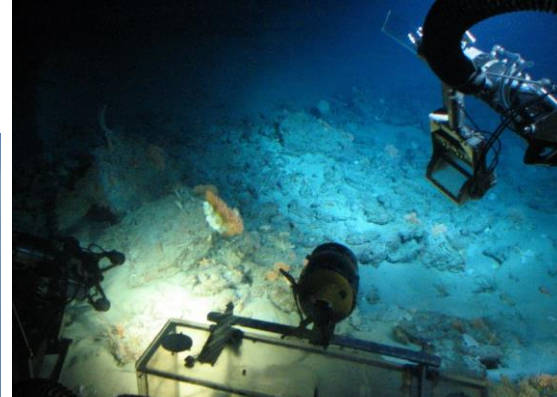
Novel food ingredients for sustainable materials



Sources of bacteria and enzymes (1)



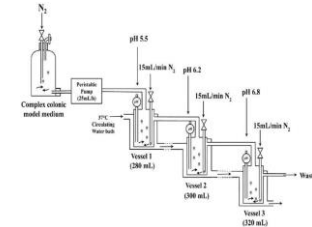
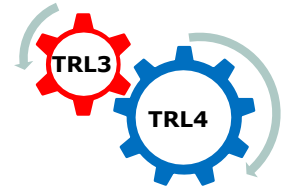
- R/V Seward Johnson
- JSL Research Submersibles
 - 1000 msw depth limit



Sources of bacteria and enzymes (2)



Production of algae-based materials



POC and
upscaling

Valorization
of side streams

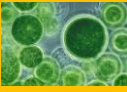
Functionalization

Prototyping

Downstream
processing

Raw material

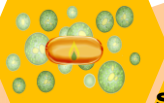
Biomass



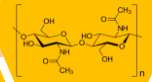
Protein



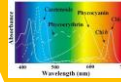
Oil



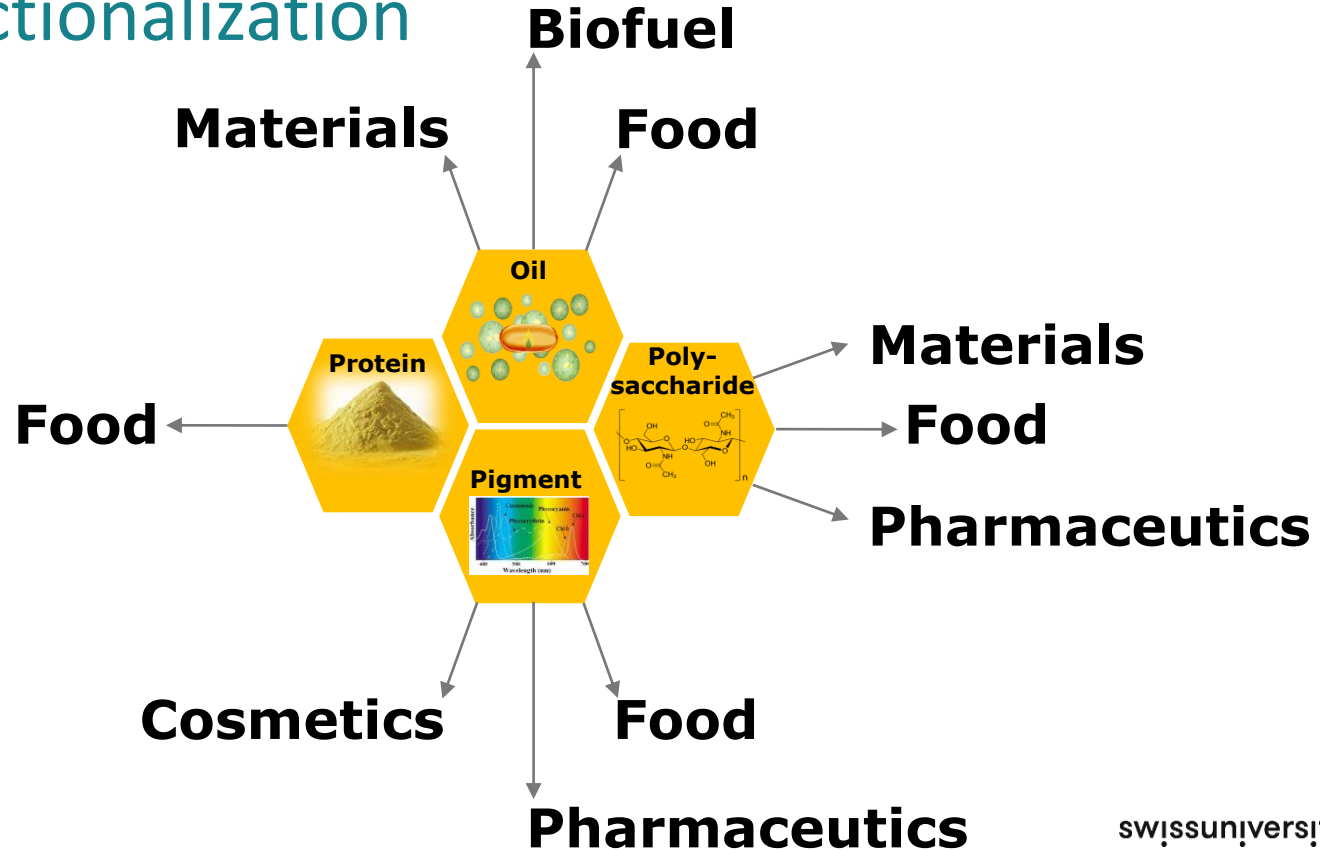
Poly-saccharide

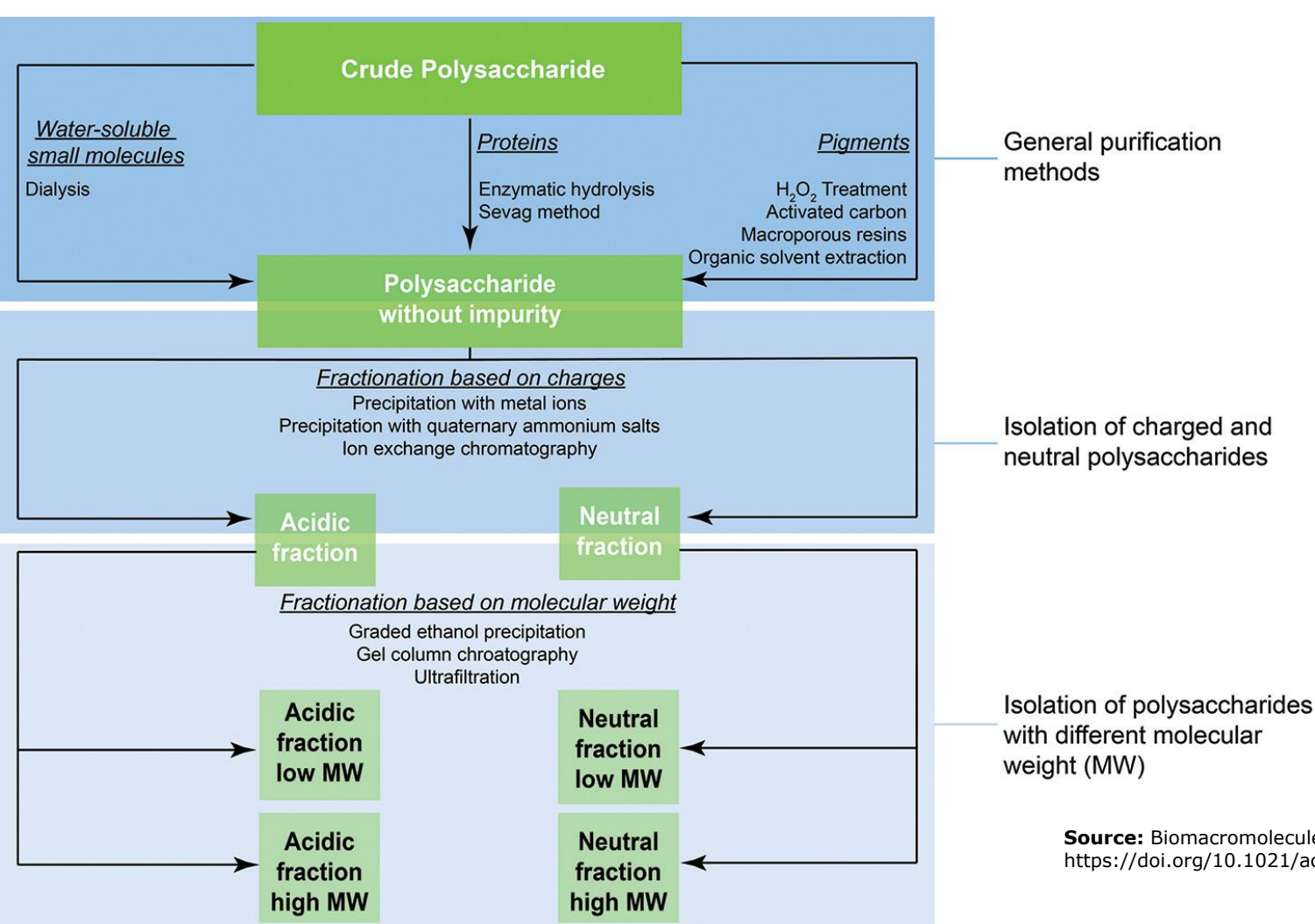


Pigment



Functionalization



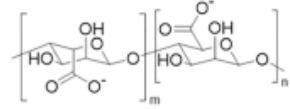
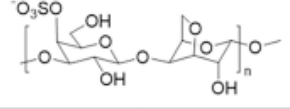
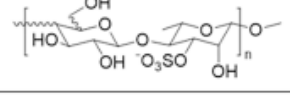
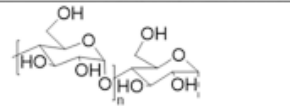
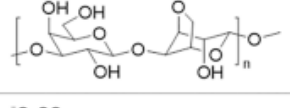
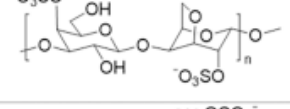
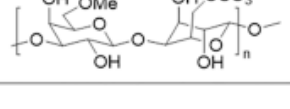
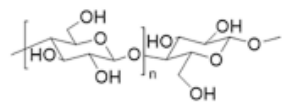


General purification methods

Isolation of charged and neutral polysaccharides

Isolation of polysaccharides with different molecular weight (MW)

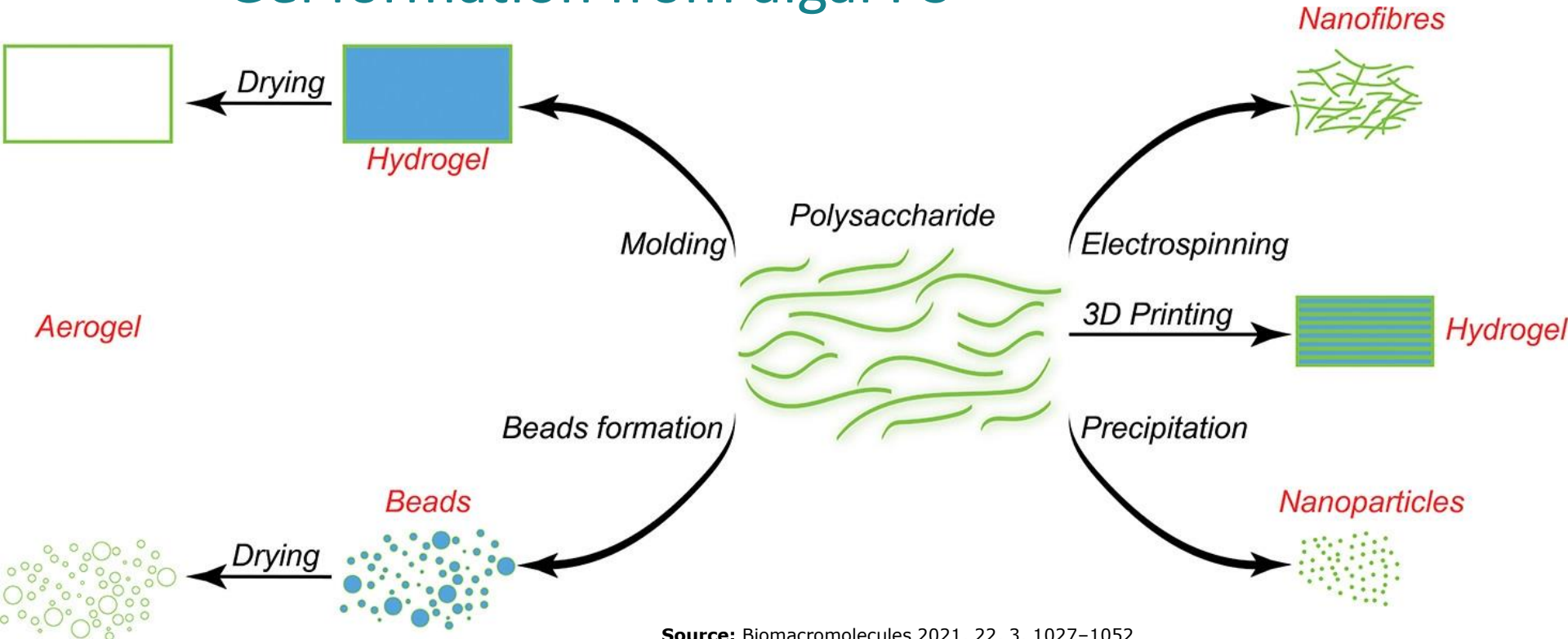
Source: Biomacromolecules 2021, 22, 3, 1027–1052
<https://doi.org/10.1021/acs.biomac.0c01406>

| | | | | |
|----------------------------------|---------------------------------|---|--|----------------------------|
| Complexation | Alginate | β -D-mannuronate α -L-guluronate |  | Ochrophyta |
| | λ -Carrageenan | β -D-galactose α -D-galactose |  | Rhodophyta |
| | Ulvan | β -D-glucuronic / α -L-iduronic acid α -L-rhamnose |  | Chlorophyta |
| Crystallization | Starch (Amylose) | α -D-glucose |  | Chlorophyta Rhodophyta |
| | | | | |
| Formation of secondary structure | Agarose | β -D-galactose 3,6 anhydro- α -L-galactose |  | Rhodophyta |
| | ι / κ -Carrageenan | β -D-galactose 3,6-anhydro- α -D-galactose |  | Rhodophyta |
| | Porphyran | β -D-galactose 3,6-anhydro- α -L-galactose |  | Prophyra |
| Colloidal assembly | Cellulose | β -D-glucose |  | Chlorophyta, ochrophyta |
| | | | | |

Algal polysaccharides

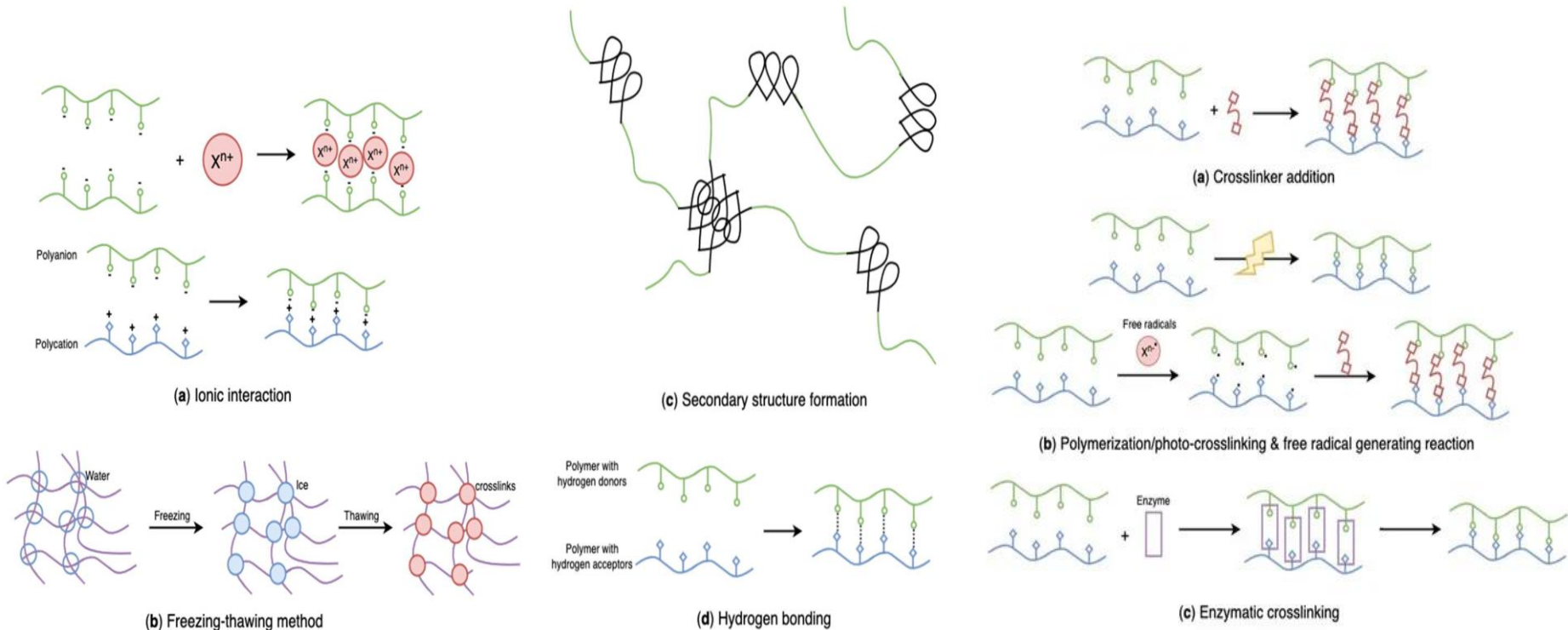
Source: Biomacromolecules 2021, 22, 3, 1027–1052
<https://doi.org/10.1021/acs.biomac.0c01406>

Gel formation from algal PS



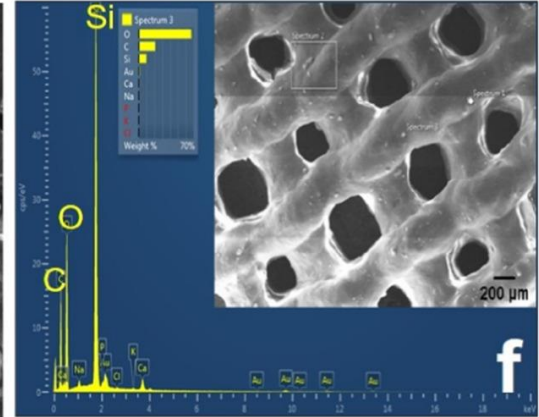
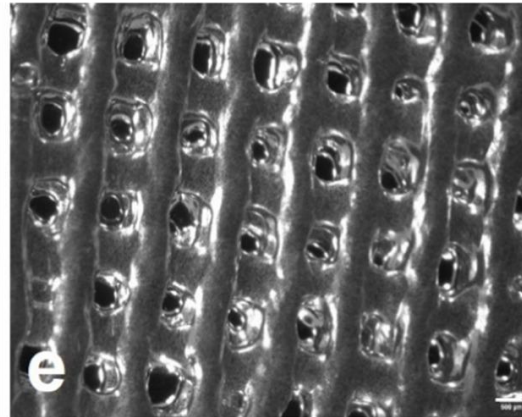
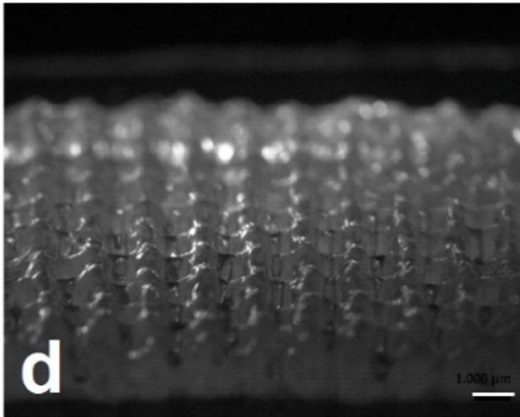
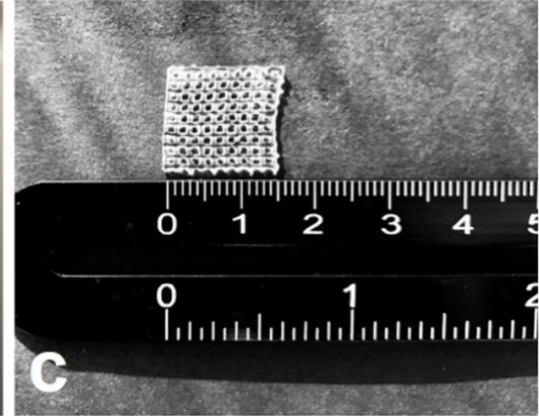
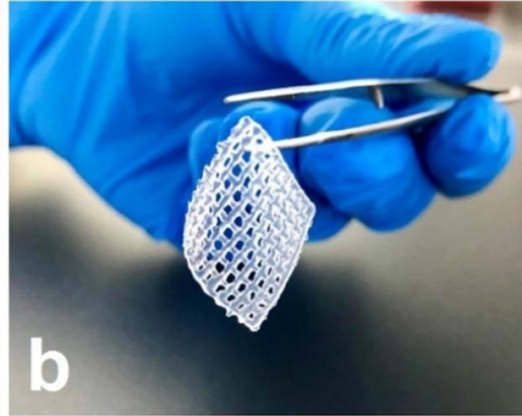
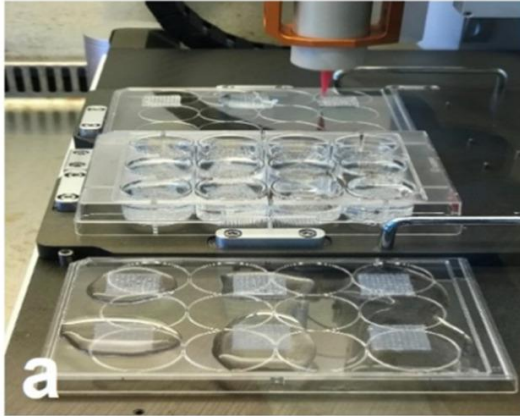
Source: Biomacromolecules 2021, 22, 3, 1027–1052
<https://doi.org/10.1021/acs.biomac.0c01406>

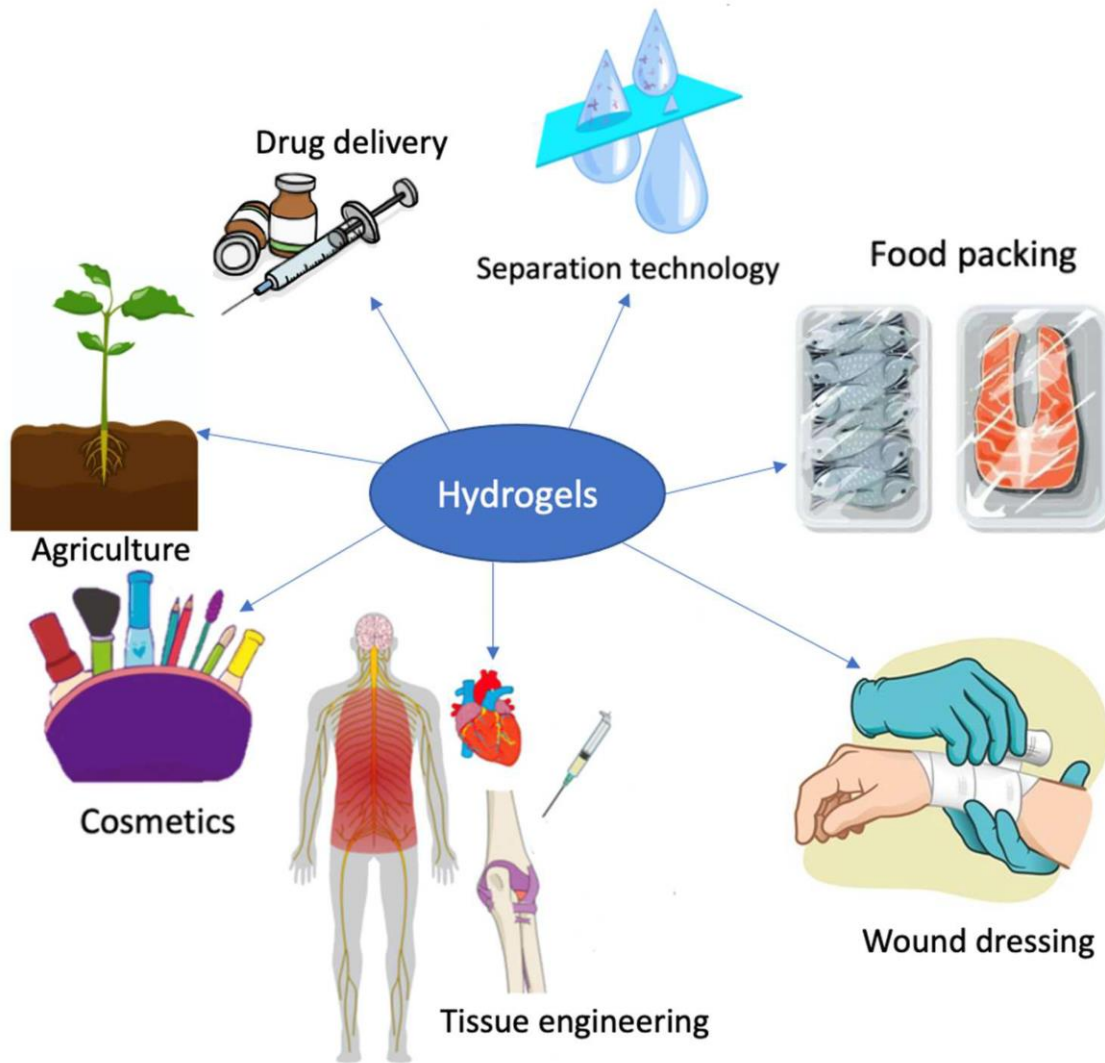
Schematic representation of physically and chemically crosslinked, algal polysaccharides-based hydrogels



Source: *Mar. Drugs* **2022**, *20*(5), 306
<https://doi.org/10.3390/md20050306>

3D printed algal hydrogel constructs





Industrial outlets for algal-based hydrogels

Source: *Mar. Drugs* **2022**, *20*(5), 306
<https://doi.org/10.3390/md20050306>

Technical
University
of Munich



Hes·so VALAIS
School of Engineering π WALLIS



WSSB
Werner Siemens-Lehrstuhl für
Synthetische Biotechnologie



swissuniversities

Algae-oil based products

Land use change



Rapeseed oil

Palm oil



Extraction



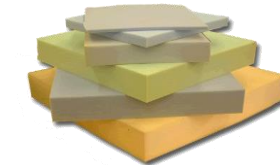
Biofuel



Food application

Food application

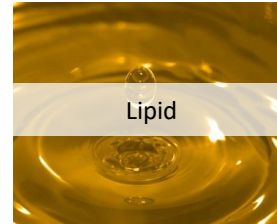
Material applications



Polymer Materials

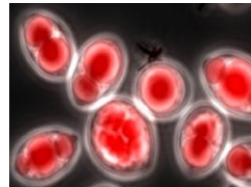


Oleochemicals

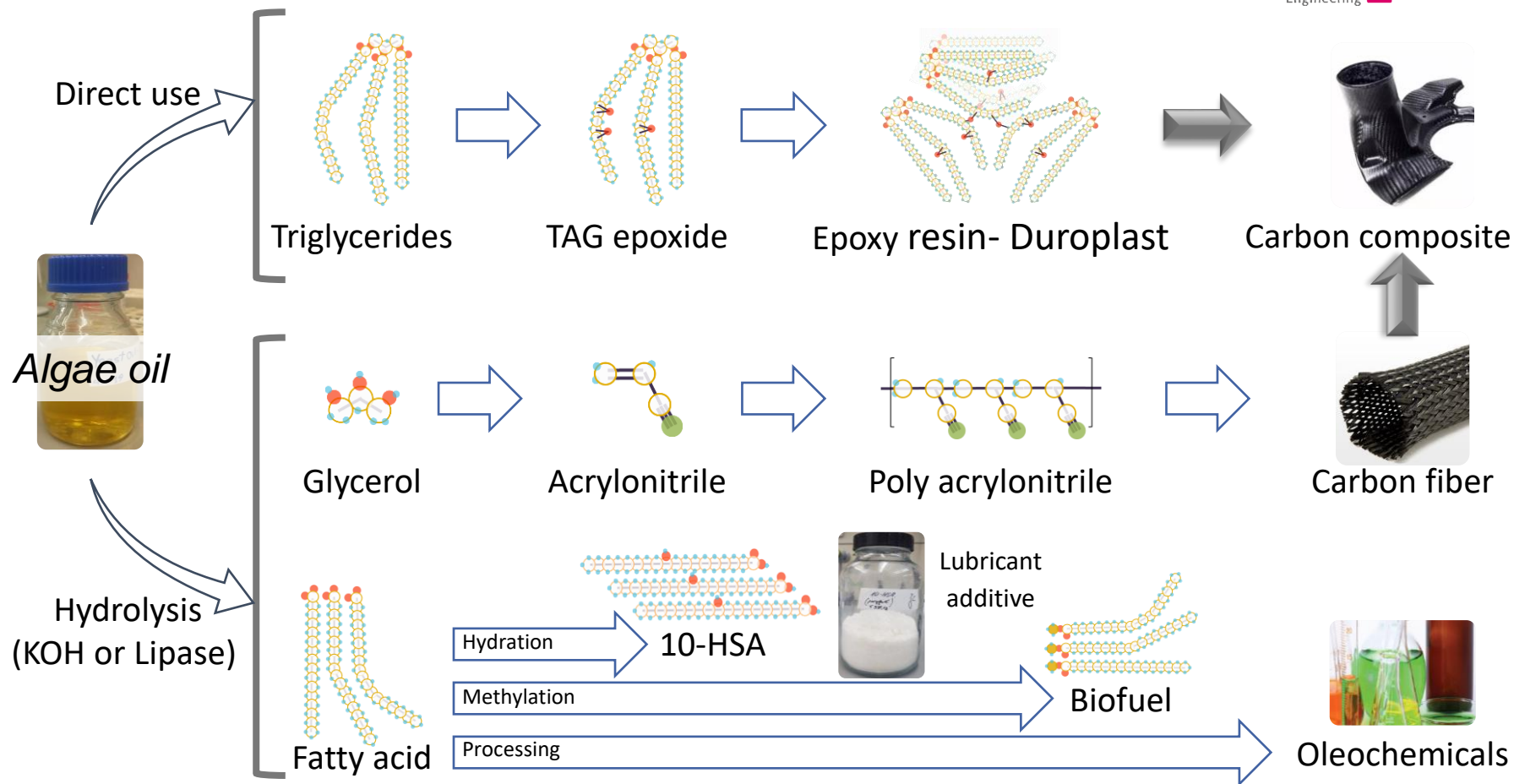


Lipid

- Waste recycling
- Sustainable oil producers
- **Microbial lipid**



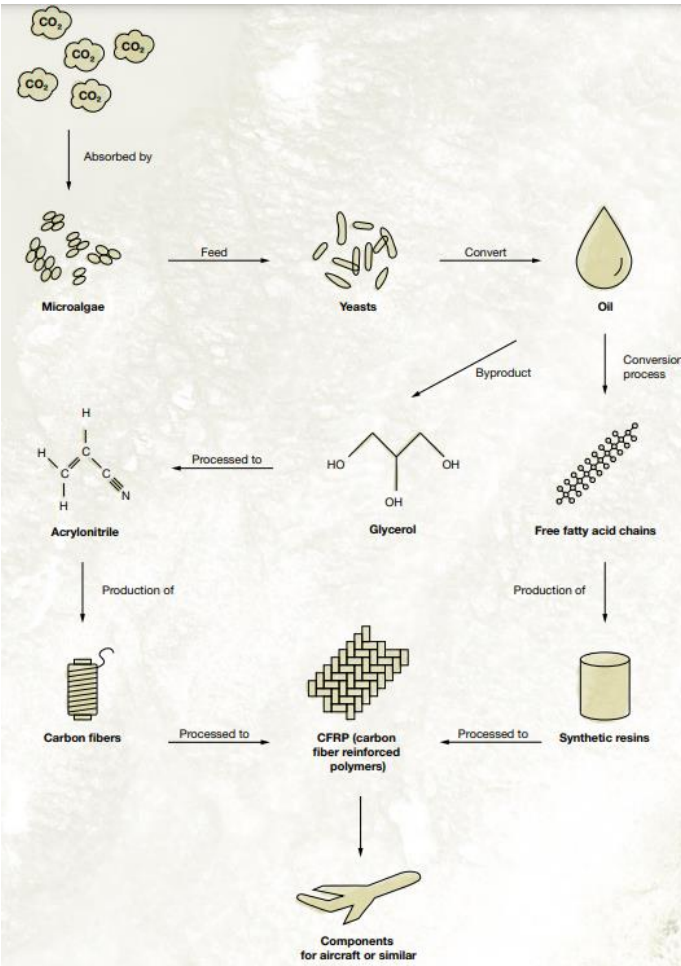
Algal lipid processing to produce carbon fiber



Carbon fiber – alternative processes

Sources:

1. <https://cseengineermag.com/innovative-materials-carbon-fibers-made-from-algae/>
2. <https://patents.justia.com/patent/20220081806>

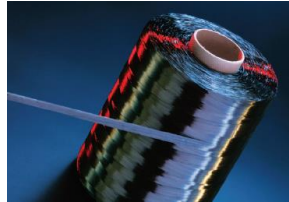


A beam made of carbon fiber reinforced granite (Mineral Carbon Composite, MCC) is load-bearing like steel, light as aluminum and extremely durable.

Existing industrial demands



supply chain from precursor fiber via CF/CFC to construction elements



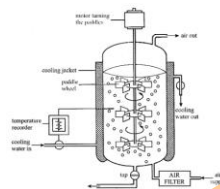
- Potential end of life CFK deposit in open cast coal mine
- Permanent CO₂ removal without toxic by products

➤ Carbon capture and removal in Gt range

Production of chitin-based materials



Benchmarking with
existing extraction
technologies



Prototyping

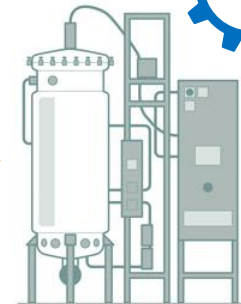
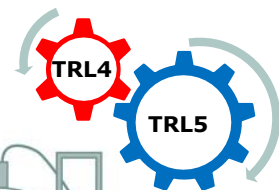
LCA, product and side-stream
characterization and management

Fermentation

Extraction through fermentation and
enzyme processing, downstream
processing, analytical product
characterization

Raw material

Pretreatment and side-
stream analysis



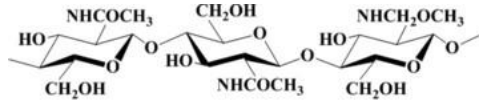
Upscaling to
technical level

Project partners: W. Brück, T. Abitbol (EPFL),
Bridge Discovery (to be resubmitted 2023)

chitin can remove some pesticides

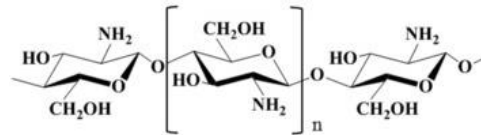
Deacetylation

Depolymerization



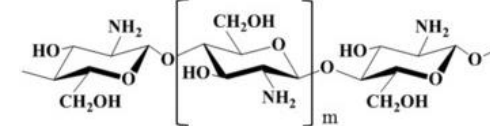
CHITIN

1. Found in insects cuticles, crustaceans shells, and fungi cell walls
2. Hard to dissolve in water, acid, and alkali solvent
3. More than 1000kDa
4. Poor Absorbability



CHITOSAN

1. Chitosan is rarely found in nature.
2. Poor water solubility, but feely dissolves in acid solvent
3. MW more than 1,000 kDa
4. Absorbability 1-3%



COS

1. Entirely dissolves in water
2. MW less than 3.9kDa
3. Absorbability almost 100%

Regardless of having various bioactivities, the water insolubilities of chitin and chitosan limit their applications in many industries

Source: (Joshi et al., 2019)

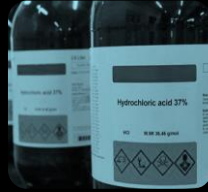


Contents of chitin and calcium carbonate in industrially important marine species

| Type | Location of chitin | Chitin (%) | CaCo3 (%) |
|---|---------------------|------------|------------|
| <u>Phylum Crustacea:</u> | | | |
| <i>Euphausia</i> sp. & <i>Meganyctiphanes</i> sp. (Krill) | cuticle/exoskeleton | 20-30 | 20-25 |
| <i>Chionoecetes</i> sp., <i>Cancer</i> sp. & <i>Carcinus</i> sp. (Crab) | cuticle/exoskeleton | 15-30 | 40-50 |
| <i>Paralithodes</i> sp. (King crab) | cuticle/exoskeleton | ~35 | 40-50 |
| <i>Callinectes</i> sp. (Blue crab) | cuticle/exoskeleton | ~14 | 40-50 |
| <i>Crangon</i> sp. & <i>Pandalus</i> sp. (Shrimp) | cuticle/exoskeleton | 17-40 | 20-30 |
| <i>Penaeus</i> sp. (Prawn) | cuticle/exoskeleton | ~40 | 20-30 |
| <i>Nephrops</i> sp. & <i>Homarus</i> sp. (Lobster) | cuticle/exoskeleton | 60-75 | 20-30 |
| <i>Lepas</i> sp. (Goose Barnacle) | shell | ~59 | 20-30 |
| <u>Phylum Mollusca:</u> | | | |
| <i>Mytilus</i> sp. & <i>Pecten</i> sp. etc. (Mussels, clams etc.) | shell | ~3 | 85-90 |
| <i>Crassostrea</i> sp. (Oyster) | shell | ~6 | 85-90 |
| <i>Loliginidae</i> sp. & <i>Ommastrephidae</i> sp. (Squid) | pen | 20-40 | negligible |

CHITOLIGOSACCHARIDES EXTRACTION METHODS

Chemical



Depolymerization with:

- Hydrochloric acid (Trombotto et al 2008)
- Nitrous acid (Mourya et al., 2011).
- Hydrogen peroxide (Hai et al., 2019)
- Phosphoric acid. (Jia, 2002)

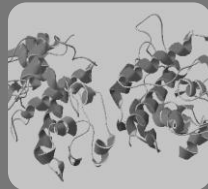
Physical



Depolymerization with :

- Microwave: (Xing et al., 2005)
- Hydrothermal and Ultrasonication (Savitri et al., 2015)

Enzymatic



• Chitinases:

- Bacterial chitinases (Kielak et al., 2013)
- Fungal chitinases (Deeba et al, 2016).
- Plant chitinases (Punja and Zhang, 1993)

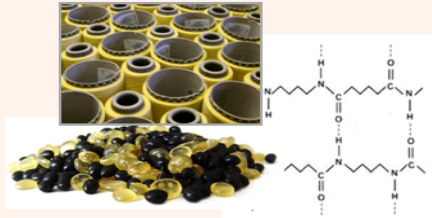
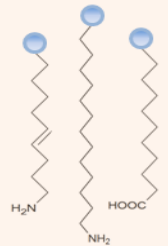
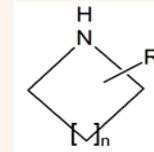
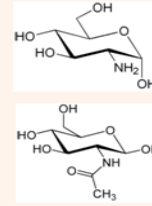
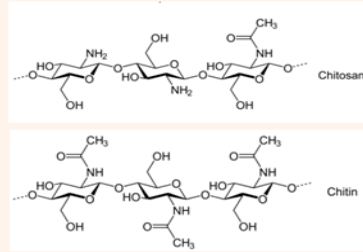


ChBio is funded by the European Commission within the Seventh Framework Programme

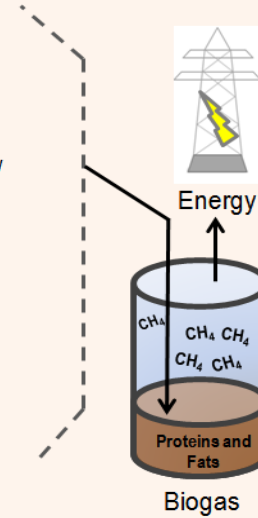
Chitin Biorefinery



Chitin-rich fishery wastes



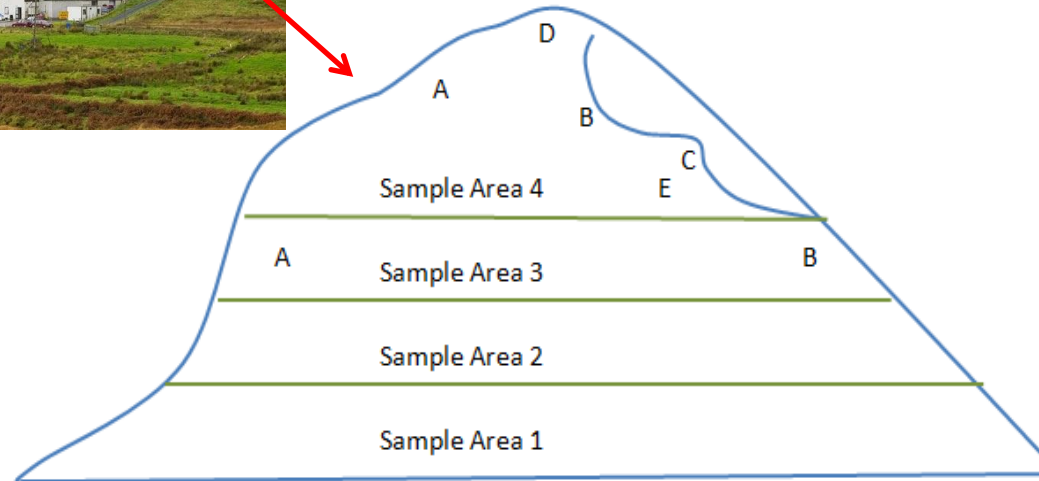
Bifunctional monomers for the chemical industry



Accumulation of marine chitin-degrading microbiota (on crab-shell waste)



Accumulation of chitin-degrading microbiota (on crab-shell waste dumping site)



Map of Sampling Site at Earagail Eisc Teoranta Ltd

Chemical treatment

Washing, drying and crushing of the shells

Alkali treatment
(1M NaOH for 1 – 72 h
at 65 – 100 °C)

Acidic treatment
(0.275 – 2.0 M HCl for
1 - 48 h at 0 – 100 °C)

Discolouration and bleaching
(organic mixture of chloroform,
methanol and water 1:2:4 at 25 °C)

Deacetylation
(NaOH/KOH -30-50 % (w/v)
at 80 - 150 °C)

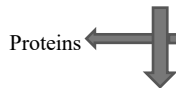
Enzymatic/chemical hydrolysis



Crustacean shellwaste from food processing



Crushed shells



Proteins

Minerals
(calcium carbonate
& calcium phosphate)

Deproteinization

Demineralization

Minerals
(calcium carbonate
& calcium phosphate)



Proteins and Pigments

Demineralization

Deproteinization

Pigments (carotenoids)

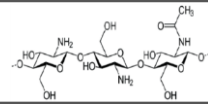
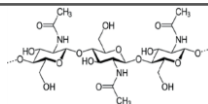
Chitin



Chitosan



Chito-oligosaccharides



Biological treatment

Washing, drying and
crushing of the shells

Organic acid (lactic acid)
producing bacteria

Protease producing bacteria

Deacetylation
(Chitin deacetylase
or lactic acid bacteria)

Hydrolysis by chitinolytic
enzymes

Preparation of cooked, minced brown crab for chitin extraction processes



Starting Material



Step 1: Hammering



Step 2: Homogenising



Step 3: Drying @55°C

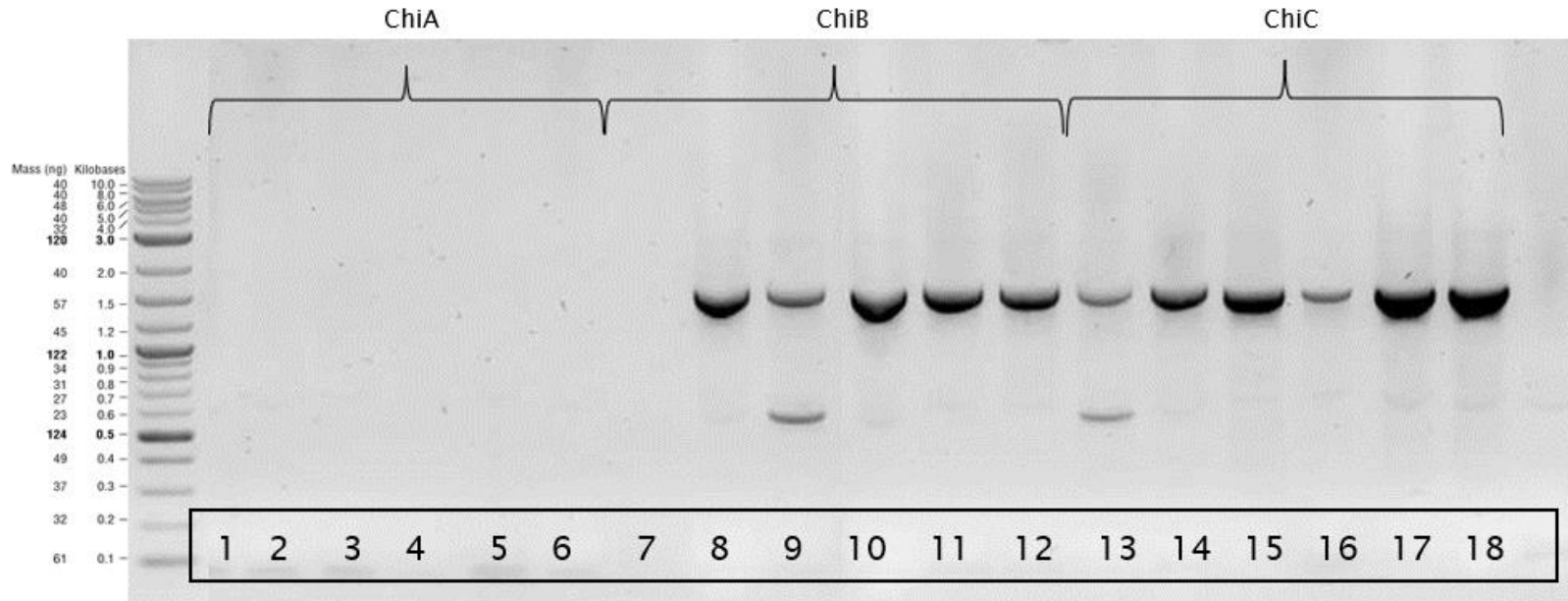


Step 4: Grinding



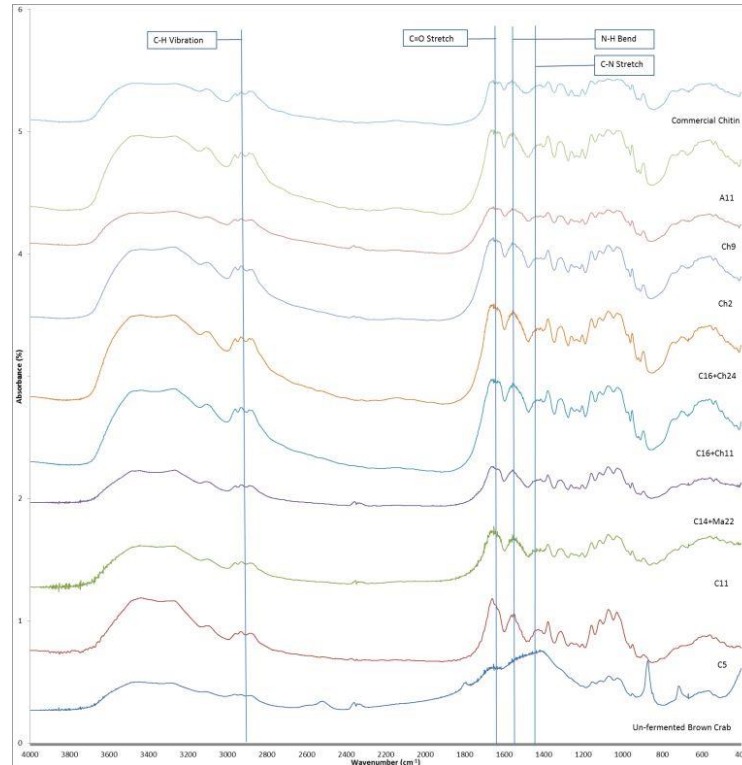
Workable raw Material

PCR products visualized on 1% agarose GEL ELECTROPHORESIS



- Presence of ChiB and ChiC genes in the three *Serratia* isolates used. ChiA is not present.

Quality assessment



← Commercial chitin

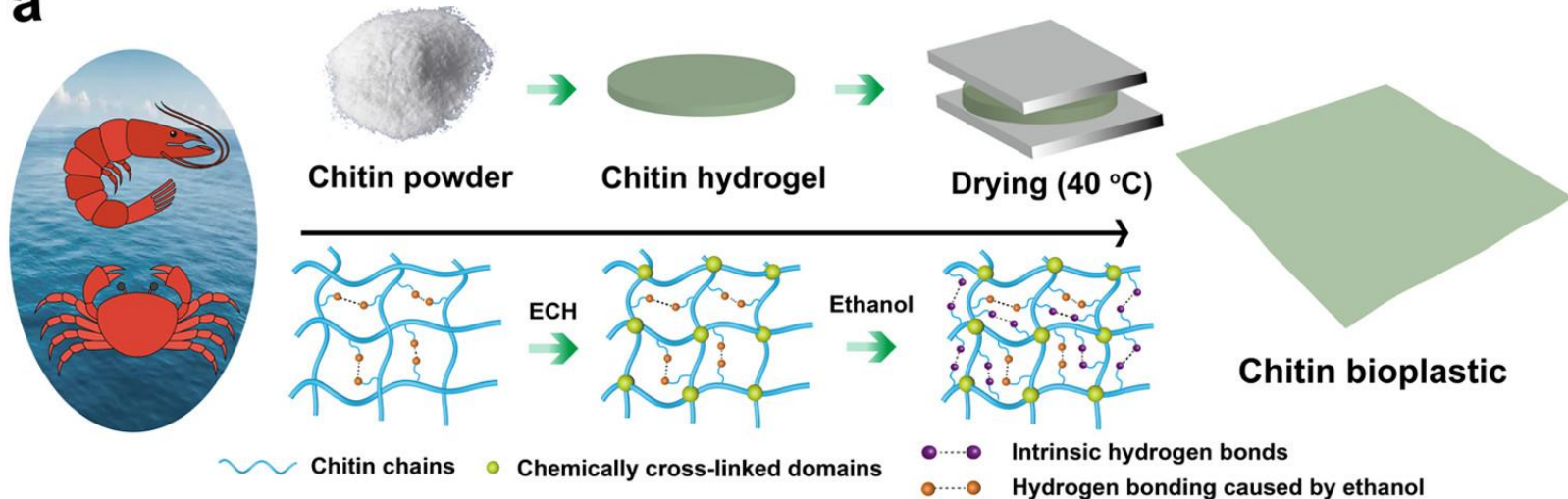
← Fermented brown crab shell

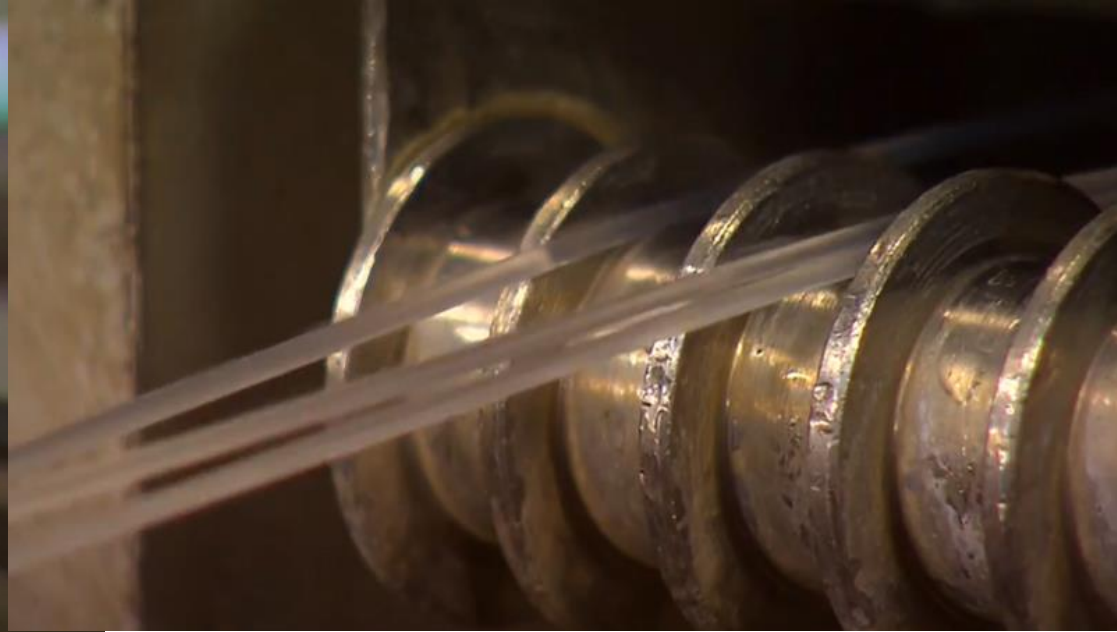
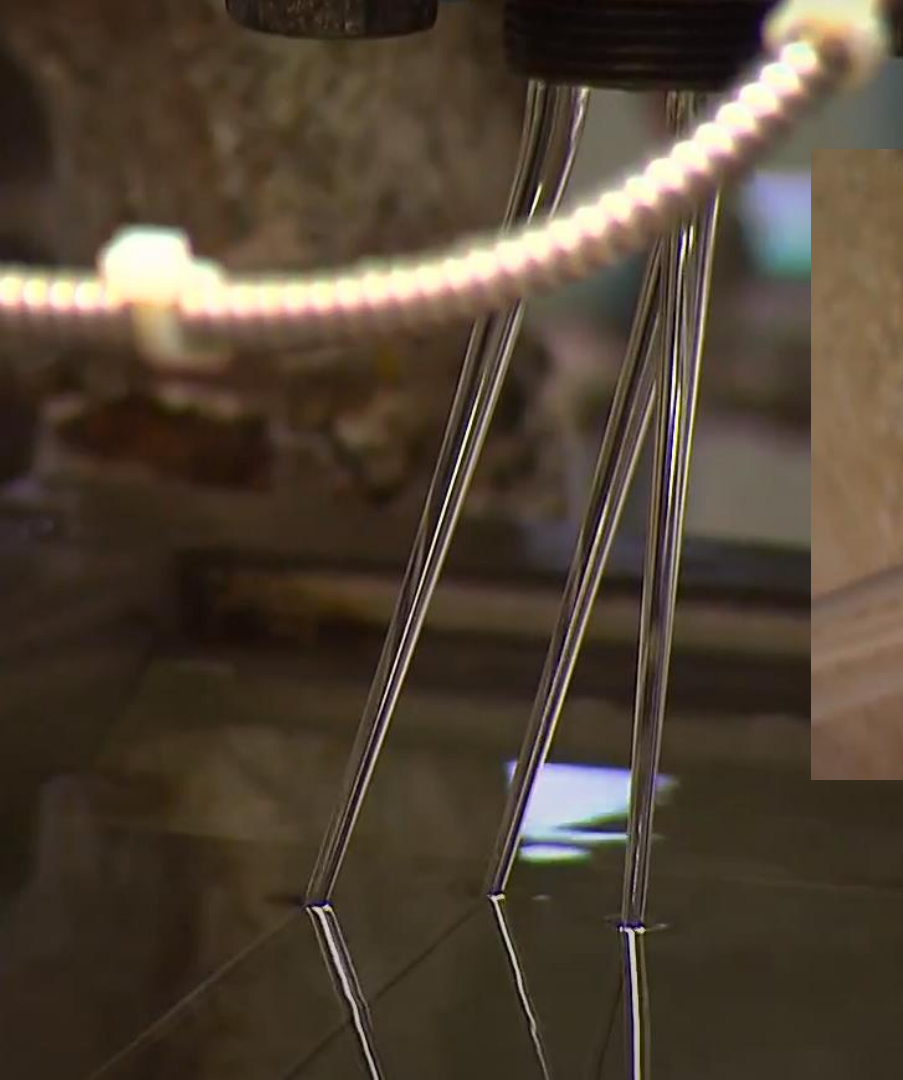
← Brown crab shell (raw)

FTIR spectra of all isolates are similar to the commercial chitin.

Production of chitin-based bioplastics

a





Bulk-produced polyamide (PA) 6.12,
containing 3% of one of the ChiBio key
monomers

Conclusions

- A circular economy for food mimics natural systems of regeneration so that waste does not exist but is instead feedstock for another cycle.
- Food side-streams can:
 - Provide additional value by creating new products.
 - Be used to revolutionize material science for the building, manufacturing, food, pharmaceutical or cosmetic industries.
 - Combat climate change
- Food side-streams have no eco-toxicology or biodiversity impact
- A Circular Food Biorefinery is a new route to an eco- and cost- efficient bioeconomy

Thank you for your attention.



Thomas Brück



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Aline Penedo



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